# GTI Diagnostic Clinic Submission Form

## Contact Information:
Phone: 519-824-4120 x 58873  
Email: turfdiag@uoguelph.ca  
Fax: 519-766-1704

## Service Summary:
- **Service option:** Diagnosis via e-mail or telephone  
  Per sample $120.00  
- Nematode extraction only  
  Per sample $50.00

## Payment Information:
Payment will be processed once sample has been processed. Info will not be kept on file.

- Cheque (enclosed with sample): Payable to “University of Guelph”
- VISA / MC #: ____________ ____________ ____________ ____________  
  Expiry ____ / ____

Name on card: ____________________________ SIGNATURE: ____________________________

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## Important: Please fill in all contact information and background details for sample. This will help ensure a quick and accurate diagnosis. Send via courier to ensure next day arrival. Instructions for taking a sample are located on the back of this form.

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## Send Sample to:
The GTI Diagnostic Clinic c/o Dr. Katerina S. Jordan  
Dept of Plant Agriculture, E. C. Bovey Building  
University of Guelph, 50 Stone Road East  
Guelph, ON N1G 2W1

## Drop off:
4224 E.C.Bovey Building (park at meters on South Ring Rd.)  
University of Guelph (intersection of Gordon St. and South Ring Rd.)  
Contact clinic to make appointment or leave on table outside lab
**SAMPLE INFORMATION SHEET**

Date symptoms first appeared*: ______________ Date sample taken*: ______________

Weather conditions at onset of symptoms (Temperature, rainfall, other)*: ________________________________

Species (if known): ________________________ Cultivar/Variety (if known): ________________________

Check the following as they pertain to your problem (information with asterisks will likely lead to a faster diagnosis)

<table>
<thead>
<tr>
<th>LOCATION*</th>
<th>ORIGIN/AGE*</th>
<th>PATTERN OF DAMAGE</th>
<th>DEGREE OF INJURY</th>
<th>SOIL CONDITIONS</th>
</tr>
</thead>
<tbody>
<tr>
<td>Green#</td>
<td>Sod</td>
<td>General</td>
<td>Light</td>
<td>pH*</td>
</tr>
<tr>
<td>Tee #</td>
<td>Seeded</td>
<td>Scattered</td>
<td>Moderate</td>
<td>Thatch levels:</td>
</tr>
<tr>
<td>Fairway#</td>
<td>Thinning</td>
<td>Severe</td>
<td>Compacted</td>
<td>Low</td>
</tr>
<tr>
<td>Lawn</td>
<td>Age*: _____</td>
<td>Rings</td>
<td>Severe</td>
<td>Moderate</td>
</tr>
<tr>
<td>Sports field</td>
<td></td>
<td>Patches</td>
<td>Rings</td>
<td>High</td>
</tr>
<tr>
<td>Sod Farm</td>
<td></td>
<td>Leaf spots</td>
<td></td>
<td>Root zone material:</td>
</tr>
</tbody>
</table>

Describe the problem in detail (i.e. symptoms, plant parts affected, distribution of symptoms):

__________________________________________________________________________

__________________________________________________________________________

Disease or disorder history of the site:

__________________________________________________________________________

Were fungicides or fertilizers applied recently? Specify type of products (s) and date (s) of application:

__________________________________________________________________________

Additional comments and specific requests:

__________________________________________________________________________

**Photos are also helpful. Please include photos or send via email to turfdiag@uoguelph.ca. Be sure to include your name and other information so we know which picture is for which sample.**
Instructions for Taking and Submitting a Sample

DISEASE/PATHOLOGY SAMPLING

1. Sample BEFORE you treat with fungicides. Fungicide application destroys signs of pathogens impeding diagnosis.
2. Sample should be 10 to 15 cm² (cup cutter size is ideal) and include foliage, thatch and at least 5 cm of roots and soil.
3. Sample should show a range of symptoms and include healthy, slightly affected and severely affected grass. Take the sample from the outside edge of a ring or patch that includes healthy and unhealthy turf. If symptoms are general, collect the sample from an area with intermediate severity. NOTE: a completely dead sample is not suitable for diagnosis.
4. Do not allow the sample to dry out or to be exposed to excessive heat or cold prior to submission.
5. Sample should be wrapped in newspaper and then in plastic and placed in a sturdy box.

NEMATODE SAMPLING

- A cup cutter can be used but will limit total sampling area and can be expensive for shipping; It is better to take multiple small extraction plugs (2cm diameter minimum) from various areas throughout affected zones
- 10 cores is the minimum – 20 to 30 is better
- Cores should be 10 cm or 4” deep
- The top 2 cm of grass, thatch and plant material can be removed from the plugs/cores

METHOD 1:

1. Take samples only from the edge of affected areas – DO NOT include dead patches or asymptomatic areas.
2. This method relies on threshold data to confirm whether the issue you are experiencing is related to nematodes or not.
3. This method does not take into account baseline populations in asymptomatic areas, and therefore produces limited information regarding nematodes on your property.

METHOD 2: (Recommended)

1. Take separate samples from both symptomatic areas and asymptomatic areas and request that each sample be assessed separately.
2. Keep these samples separate from each other as they will be considered two separate samples.
3. This method is more expensive but will produce information on your baseline levels in asymptomatic areas as well as threshold population data in the affected areas.

METHOD 3:

1. Take samples systematically throughout entire area (ie. every 15 feet) including both affected and non-affected areas and combine the cores into one sample.
2. This method is useful for determining overall population levels but is not useful in associating symptoms with nematode populations.

WHEN SUBMITTING A SAMPLE:

1. Fill out the submission form as completely as possible and include it with the sample
2. Do not send samples by regular mail or over a weekend! They must be sent so that they arrive NEXT DAY.
3. Photos are not necessary but do help with the diagnostic process.

*You do not need to include this sheet with your submission*